Bioconversion of dietary provitamin A carotenoids to vitamin A in humans

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ABSTRACT
Recent progress in the measurement of the bioconversion of dietary provitamin A carotenoids to vitamin A is reviewed in this article. Methods to assess the bioavailability and bioconversion of provitamin A carotenoids have advanced significantly in the past 10 y, specifically through the use of stable isotope methodology, which includes the use of labeled plant foods. The effects of the food matrix on the bioconversion of provitamin A carotenoids to vitamin A, dietary fat effects, and the effect of genotype on the absorption and metabolism of β-carotene have been reported recently. A summary of the major human studies that determined conversion factors for dietary β-carotene to retinol is presented here, and these data show that the conversion efficiency of dietary β-carotene to retinol is in the range of 3.6–28:1 by weight. There is a wide variation in conversion factors reported not only between different studies but also between individuals in a particular study. These findings show that the vitamin A value of individual plant foods rich in provitamin A carotenoids may vary significantly and need further investigation. *Am J Clin Nutr* 2010;91(suppl):1468S–73S.

INTRODUCTION
Dietary provitamin A carotenoids are a major source of our vitamin A needs. Vitamin A is an essential vitamin for the promotion of general growth, maintenance of visual function, regulation of differentiation of epithelial tissues, and embryonic development (1). Vitamin A can be obtained from food, either as preformed vitamin A in animal products, such as eggs and dairy products, or as provitamin A carotenoids, mainly β-carotene in plant products, such as green leafy and yellow-colored vegetables and orange-colored fruit.

In Western societies, the provitamin A carotenoids derived from plants provide <30% of daily vitamin A intake, whereas preformed vitamin A derived from animal products provides >70% daily vitamin A intake (2). In contrast, in developing countries, provitamin A carotenoids in vegetables and fruit provide >70% of daily vitamin A intakes (3).

Epidemiologic data have shown that diets rich in carotenoid-containing foods are associated with decreased risk of certain types of chronic diseases, such as cancer (4), cardiovascular disease (5), age-related macular degeneration (6, 7), and cataracts (8, 9). The disease-preventing activity of β-carotene and other provitamin A carotenoids could be ascribed either to their conversion into retinoid or to their activity as intact molecules. The results of several human intervention studies, however, indicate that high-dose supplementation with β-carotene, either alone (10) or with vitamin E (11) or vitamin A (12), does not decrease the risk of cancer or cardiovascular disease, and might even be harmful to smokers or former asbestos workers. Thus, it may be that β-carotene and other carotenoids promote health when taken at physiologic amounts in foods, but have adverse properties when given in high doses and under highly oxidative conditions. Furthermore, the health benefits of food β-carotene consumed at a physiologic amount as an intact molecule and/or its cleavage product (vitamin A) should be investigated to determine the relative importance of each potentially favorable property in various populations. The conversion of dietary β-carotene to vitamin A may relate to preformed vitamin A in the diet (13); that is, the conversion may be less efficient when vitamin A has been provided from other dietary sources. The issue of the efficiency of conversion of provitamin A carotenoids into retinol and other retinoids is therefore of interest for us to better understand the nutritional value of dietary provitamin A carotenoids.

It is well established that after an oral dose of β-carotene, both intact β-carotene and its metabolite retinol can be found in the circulation. In humans, conversion of β-carotene into vitamin A takes place predominantly in the intestine and less so in other tissues. The ratio of the amount of β-carotene given in an oral dose to the amount of vitamin A derived from this β-carotene dose is defined as the β-carotene-to-vitamin-A conversion factor or β-carotene equivalent to vitamin A.

For a healthy population, the major factors that affect the bioavailability of food carotenoids and the bioconversion of food...
provitamin A carotenoids to vitamin A in humans are food matrices, food preparation, and the fat content of a meal (14). Recent studies reported that the conversion efficiency of dietary β-carotene is in the range of 10 to 28:1 by weight (15–19). These data indicated that the bioconversion of β-carotene to vitamin A was not as efficient as expected, and, as a result, the Food and Nutrition Board recently revised the estimated efficiency factor for the conversion of dietary β-carotene to vitamin A from 6:1 by weight (20) to the new value of 12:1 by weight (21). However, this new conversion ratio must be regarded as temporary and could well change, as more data become available.

On the other hand, preformed vitamin A from animal origins or from supplements can be absorbed and stored in the human body very effectively. A recent report (22) showed that among women who did not take supplemental vitamin A, retinol from food was very effectively absorbed and stored in the human body as temporary and could well change, as more data become available. Different methods have been developed and used to determine these bioconversion factors in various populations with different dietary intakes. For populations with normally low vitamin A intakes, DRD can be used to measure changes of total body stores of vitamin A. A DRD method was used in a study of children with marginal-to-normal vitamin A status, who participated in a food-based intervention with either green-yellow vegetables or light-colored vegetables with low carotene content (18). The serum carotenoid concentrations of children fed green-yellow vegetables increased, whereas the serum concentration of vitamin A did not change. In contrast, the DRD tests carried out before and after the vegetable intervention showed that the body stores of vitamin A were stable in the group fed green-yellow vegetables (19, 20). Instead, DRD can be used to measure changes of total body stores of vitamin A. A DRD method was used in a study of children with marginal-to-normal vitamin A status, who participated in a food-based intervention with either green-yellow vegetables or light-colored vegetables with low carotene content (18). The serum carotenoid concentrations of children fed green-yellow vegetables increased, whereas the serum concentration of vitamin A did not change. In contrast, the DRD tests carried out before and after the vegetable intervention showed that the body stores of vitamin A were stable in the group fed green-yellow vegetables (19, 20). Instead, DRD can be used to measure changes of total body stores of vitamin A.

Changes in whole-body stores of vitamin A (paired deuterated retinol dilution)

Advances in isotope technology have facilitated the measurement of changes in body stores of vitamin A after the feeding of dietary provitamin A carotenoids; specifically, paired deuterated-retinol-dilution (DRD) tests can be used to measure the conversion efficiency in food-based intervention studies. For populations with marginal-to-normal vitamin A status, changes in serum retinol concentrations may not be a sensitive indicator of vitamin A status. Instead, DRD can be used to measure changes of total body stores of vitamin A. A DRD method was used in a study of children with marginal-to-normal vitamin A status, who participated in a food-based intervention with either green-yellow vegetables or light-colored vegetables with low carotene content (18). The serum carotenoid concentrations of children fed green-yellow vegetables increased, whereas the serum concentration of vitamin A did not change. In contrast, the DRD tests carried out before and after the vegetable intervention showed that the body stores of vitamin A were stable in the group fed green-yellow vegetables (19, 20). Instead, DRD can be used to measure changes of total body stores of vitamin A.
was seen in the children fed light-colored vegetables that contained little \( \beta \)-carotene, but 275 mg \( \beta \)-carotene from green-yellow vegetables prevented this loss. From this paired DRD test, it was calculated that 27 \( \mu \)g \( \beta \)-carotene from vegetables was equivalent to 1 \( \mu \)g retinol. This conversion factor is similar to that reported in another study, which measured changes in serum vitamin A concentration after consumption of carotenoids from vegetables (17).

The paired DRD technique has also been used (15) to measure change in the vitamin A pool size after 60-d supplementation with 750 RE/d as either retinyl palmitate, \( \beta \)-carotene, sweet potato, or Indian spinach, compared with a control that contained no retinol or carotene (\( n = 14 \)/group). Vitamin A equivalency factors of 6:1 for \( \beta \)-carotene in oil, 10:1 for \( \beta \)-carotene in Indian spinach, and 13:1 for \( \beta \)-carotene in sweet potato were determined. A recent study used a mixed-vegetable intervention and the paired DRD test to measure changes in vitamin A pool size (27). The results showed that the conversion factors were better than 12:1 for \( \beta \)-carotene and 24:1 for other provitamin A carotenoids.

**CONSUMPTION OF INTRINSICALLY LABELED DIETARY PROVITAMIN A CAROTENOIDS**

To achieve an accurate assessment of carotenoid bioabsorption and a subsequent vitamin A value from a food source, food material is required in which the carotenoids have been endogenously or intrinsically labeled with a low-abundance stable isotope. Plant carotenoids can be intrinsically labeled either through the addition of a carbon-stable isotope presented to the roots in the form of heavy water, \( 2 \H_2\mathrm{O} \), as a gas atmosphere as \( 1\H^1\mathrm{CO}_2 \) or through the addition of a hydrogen-stable isotope presented to the roots in the form of heavy water, \( 2\H_2\mathrm{O} \). For \( 2\H_2\mathrm{O} \), plants can be labeled via hydroponic growth (28) on a nutrient solution composed of a fixed \( 2\H_2\mathrm{O} \) percentage to achieve the labeling of the carotenoids. This allows presentation of the carotenoids in their normal cellular compartments, and the isotopic label enables identification of those serum carotenoids (or derived retinol), which come from the specific food in question. The deuteriation of intrinsically labeled plant \( \H_2\beta \)-carotene has been shown to be distributed randomly throughout the carotenoid molecules (29).

We have developed an isotope reference method to quantitatively determine the retinol equivalence of \( \beta \)-carotene (either synthetic pure \( \beta \)-carotene or \( \beta \)-carotene contained in a food). In the isotope reference method, through the use of a known amount of labeled vitamin A, such as retinol acetate-\( 8\H \), as a reference and the comparison of the amount of retinol formed in vivo from a vitamin A precursor (eg, \( \H_2\beta \)-carotene), we may determine quantitatively the vitamin A value of the vitamin A precursor, as shown in Figure 1. Therefore, the retinol equivalence of the \( \H_2\beta \)-carotene dose = \((\text{AUC}_{\H_2\beta \text{retinol}}/\text{AUC}_{\H_2\beta \text{retinol}}) \times \text{Dose}_{\H_2\beta \text{retinol}} \times \text{retinyl acetate} \), where AUC is the area under the curve of the circulating tracer measured compared with time. For example, in the serum of a volunteer, if, 21 d after a 6-\( \mu \)g \( \H_2\beta \)-carotene oral dose and a 3-\( \mu \)g \( \H_2\beta \)-carotene oral dose, the area under the \( \H_2\beta \)-retinol curve (derived from the \( \H_2\beta \)-carotene) is measured as 10 units and the area under the \( \H_2\beta \)-retinol curve (from the \( \H_2\beta \)-retinol acetate) is measured as 10 units, we can say that 6 mg of synthetic \( \beta \)-carotene is nutritionally equivalent to 3 mg retinyl acetate. That is, we will be able to define the vitamin A activity of \( \beta \)-carotene or a food that contains \( \beta \)-carotene, as long as it is labeled properly. Such an isotope reference method (with a known amount of \( \H_2\beta \)-retinyl acetate as the reference material) can be used to define the vitamin A activity of various vitamin A precursors, synthetic \( \beta \)-carotene (30), or provitamin A carotenoids in vegetables or other plants in humans.

This method has been used to determine the vitamin A value of endogenously labeled spinach and carrot carotenoids (19). For these experiments on carotenoid absorbability and conversion, we recruited 7 healthy male adults who had normal vitamin A status and were nonsmokers. They were given pureed and cooked spinach (\( n = 14 \), 7 men and 7 women) and pureed and cooked carrot (\( n = 7 \) men) grown hydroponically in 25 atom % \( \H_2\mathrm{O} \). With the use of 3.0 mg labeled retinyl acetate as a reference dose, we observed that provitamin A carotenoids in carrots have greater vitamin A potency than those in spinach. The 300 g labeled spinach and 100 g labeled carrots each contained \( \approx 0.11 \) mg (all-trans)-\( \beta \)-carotene, and it was assumed, as usual, that \( \zeta \)-carotene and (cis)-\( \beta \)-carotene, which were also present, had half the activity of (all-trans)-\( \beta \)-carotene (21). The retinol equivalences were determined to be 21 \( \mu \)g spinach \( \beta \)-carotene or 15 \( \mu \)g carrot \( \beta \)-carotene to 1 \( \mu \)g retinol.

With a similar approach, dried *Spirulina* powder was studied in humans. *Spirulina* is an alga rich in \( \beta \)-carotene. With the use of hydroponically grown and intrinsically deuterium-labeled *Spirulina*, and labeled retinyl acetate as a reference dose, a study was conducted in Chinese adults (\( n = 10 \) men) with normal vitamin A status (31). The volunteers (average age, 48 y) each took 5 g dried *Spirulina* powder that contained 4.3 mg \( \beta \)-carotene. When compared with a reference dose of 2.0 mg \( 13\C_{10} \) retinyl acetate in oil administered in a capsule, 4.5 mg *Spirulina* \( \beta \)-carotene provided 1 mg retinol.

Very recently, a human study that used Golden Rice \( \beta \)-carotene was reported (32): 65−98 g (130−200 g cooked weight) of hydroponically grown Golden Rice that contained 0.99−1.53 mg of \( \beta \)-carotene was given to 5 healthy volunteers (3 women and 2 men) in the United States. With the use of the isotope reference method, in comparison with the \( 13\C_{10} \) retinyl acetate reference dose, Golden Rice \( \beta \)-carotene provided 0.24−0.94 mg retinol. Thus, the conversion factor of Golden Rice \( \beta \)-carotene to retinol is 3.6:1 with a range of 1.6 to 6.4:1 by weight. Therefore,
β-carotene derived from Golden Rice is effectively converted to vitamin A in humans.

OTHER METHODS

In the 1960s, a few studies were carried out in humans that investigated the vitamin A activity of β-carotene with the use of radioisotope-labeled β-carotene. Studies by Goodman et al (33) and Blomstrand and Werner (34) have provided most of our knowledge about how humans absorb and metabolize β-carotene. Such radioisotope methods can no longer be used for ethical reasons.

The measurement of β-carotene and retinyl esters in postprandial chylomicron fractions after the feeding of food rich in provitamin A carotenoids was developed to study a single dose of β-carotene. Postprandial chylomicron response curves of β-carotene and retinyl esters in blood were measured after a single dose of β-carotene supplement in oil or from vegetables (35–37). In these studies, triacylglycerol-rich lipoproteins (TRLs) with density <1.006 g/mL were separated and analyzed to evaluate the absorption efficiency of β-carotene (intact and, after central cleavage, as retinyl palmitate). The efficiency of absorption of β-carotene by each subject was calculated by measurement of the areas under the curve (AUC, nmol-h/L) of β-carotene and retinyl ester concentrations in postprandial TRL fractions collected hourly. These curves were compared with hypothetical AUC after an intravenous dose of the same amount of β-carotene, on the assumption that the β-carotene disappearance follows a first-order elimination from blood with a chylomicron remnant half-life of 11.5 min (35). To compensate for the variability of TRL recovery, deuterium-labeled vitamin A was used (38) as an extrinsic standard. A subject was given raw carrots that contained 9.8 μmol (5 mg) β-carotene and 5.2 μmol x-carotene (2.8 mg), together with 7 μmol (2 mg) [2H4]-retinyl acetate, and the concentrations of β-carotene, x-carotene, and labeled and unlabeled retinyl esters in the TRL were measured at various time points up to 7 h. With the assumption that absorption of labeled retinyl acetate was ~80% of the dose, it was calculated that 0.8 μmol of the carrot β-carotene was absorbed intact and that 1.5 μmol of unlabeled retinyl esters were formed from the carrot dose. The mass equivalency of carrot β-carotene to vitamin A was, therefore, 13:1 by weight (without consideration of the contribution from 5.2 μmol of x-carotene to vitamin A). If the contribution of x-carotene is considered, the ratio is higher (16:1), with the assumption that x-carotene has half the activity of β-carotene.

FACTORS THAT AFFECT THE BIOCONVERSION OF β-CAROTENE

Review articles that have evaluated the factors that affect the conversion of β-carotene to vitamin A have been published by Castenmiller and West in 1998 (39), van het Hof et al in 2000 (40), and Yeum and Russell in 2002 (41). For bioconversion of dietary β-carotene, the major factors are food matrix, food processing, and fat in the diet. As mentioned above, even though the spinach and carrots were both pureed and cooked, the in vivo study showed that spinach β-carotene had a conversion factor of 21:1, whereas carrot β-carotene had a conversion factor of 15:1. This was due to differences in the food matrix: β-carotene in spinach leaves is in the form of pigment proteins located in chloroplasts, whereas in carrots the β-carotene is in a crystal form in chromoplasts (42). In addition, the efficient conversion of Spirulina β-carotene may be due to its simple cell structure, which is composed of protein and peptidoglycans that are digested easily (43). Similarly, rice has a simple and easily digestible food matrix, which allows for a high bioavailability and bioconversion of β-carotene to vitamin A.

Intake of β-carotene at different dosage amounts will affect the conversion efficiency of β-carotene when given as β-carotene oil capsules (44). In this study a subject who took 6 mg β-carotene in oil capsules showed a conversion factor of 3.8:1, whereas the same subject who took a dose of 126 mg β-carotene in oil showed a conversion factor of 55:1 by weight. It is well known that dietary fat is a critical factor that affects bioavailability and bioconversion of β-carotene. However, the amount of fat needed in the diet to facilitate the absorption of carotenoids should be studied. A recent study on the influence of amounts of dietary fat on the bioavailability and bioconversion of provitamin A carotenoids in yellow and green leafy vegetable meals showed that carotenoid-rich yellow and green leafy vegetables need a certain minimum amount of fat (2.4 g fat/meal) to ensure the absorption of fat-soluble provitamin A carotenoids and to improve vitamin A nutritional status (27).

Our study also showed that there is a correlation of the β-carotene-to-vitamin A conversion factor with the BMI in individual subjects (30). That is, conversion factors of the subjects who received synthetic β-carotene were significantly correlated with BMI. This suggests that subjects with more body fat have a lower capability to convert β-carotene to vitamin A.

We have observed large variations in the bioconversion of dietary β-carotene to vitamin A, which may be related to the genetic characteristics of the subjects. Because the enzyme responsible for β-carotene conversion into retinol is β-carotene 15,15′-monooxygenase (BCMO1), genetic polymorphisms in the BCMO1 gene may contribute to the poor converter phenotype. Very recently, it was reported (45) that 2 common nonsynonymous single nucleotide polymorphisms (R267S and A379V) had been identified and in vitro biochemical characterization of the recombinant 267S + 379V double mutant showed a decreased catalytic activity of BCMO1 by 57% (P < 0.001). A further assessment of the responsiveness to a pharmacologic dose of β-carotene in female volunteers confirmed that carriers of the 379V and 267S + 379V variant alleles showed a 69% decrease in their ability to convert β-carotene and a 240% increase in fasting plasma β-carotene concentration. Therefore, there is genetic variability in β-carotene metabolism. This may provide an explanation for the molecular basis of the poor converter phenotype within the population. Multiple single nucleotide polymorphisms and genes that might influence β-carotene status warrant further study.

CONCLUSIONS

Provitamin A carotenoids from various foods have been shown to have an almost 8-fold difference in β-carotene conversion factors (on a weight basis) that ranged from 3.6:1 to 28:1 with Golden Rice and leafy vegetables, respectively (Table 1), and thus have different values in terms of vitamin A nutrition. The major factor that affects the vitamin A value of plant provitamin
A carotenoids is the food matrix. Stable isotope labeling has provided much of the technology to study the bioconversion of vegetable provitamin A carotenoids to vitamin A at a dietary level in various populations. Dietary provitamin-A-carotenoids-to-vitamin-A conversion factors can be used in the development of dietary guidelines in well-nourished populations, and ultimately used to help combat vitamin A deficiency worldwide. Future studies on various plant foods, which include staple foods rich in provitamin A carotenoids, will be needed to both discover and evaluate vitamin A–rich sources of plant foods.

It should be remembered that human subjects may have different abilities to convert provitamin A carotenoids to vitamin A. These differences in conversion efficiency may be due to the genetic variability in β-carotene metabolism of individual human subjects. Therefore, provitamin A carotenoids might not be a good vitamin A source for those subjects of the poor converter phenotype.

The author had no conflict of interest and gained no financial benefit from writing this manuscript.

REFERENCES


TABLE 1
Summary of studies to determine a conversion factor for β-carotene in food sources to vitamin A

<table>
<thead>
<tr>
<th>Food matrix</th>
<th>Method</th>
<th>Dose</th>
<th>Conversion factor (by weight)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fruit, n = 49; vegetables, n = 45; retinol-rich foods, n = 48 (ages 7–11 y)</td>
<td>Changes of serum retinol concentration in vitamin A-deficient (~0.7 μmol/L) anemic schoolchildren</td>
<td>Fruit: 509 RE/d; vegetables: 684 RE/d; vitamin A-rich foods: 556 RE/d</td>
<td>12:1</td>
<td>16</td>
</tr>
<tr>
<td>Green/yellow vegetables, n = 10; light-colored vegetables, n = 8</td>
<td>Total body stores of vitamin A before and after the vegetable intervention in schoolchildren (aged 5.3–6.9 y) with normal or marginal vitamin A status</td>
<td>Green/yellow vegetables (206 μg calculated trans-β-carotene) to prevent the decrease of 7.7 μg in liver stores</td>
<td>27:1</td>
<td>18</td>
</tr>
<tr>
<td>Sweet potato, Indian spinach, beta-carotene capsule, or retinyl palmitate; all, n = 14</td>
<td>Mean changes of total body stores of vitamin A before and after a 60-d intervention in adult men compared with the mean changes in the retinyl palmitate group</td>
<td>Sweet potato: 750 μg RE; Indian spinach: 750 μg RE; β-carotene capsule: 750 μg RE; retinyl palmitate: 750 μg RE</td>
<td>13:1</td>
<td>15</td>
</tr>
<tr>
<td>[3H]-Labeled spinach, and vitamin A in oil capsule, n = 14</td>
<td>Comparison of AUC responses to the spinach and the vitamin A reference dose in adults</td>
<td>Calculated 11 mg trans-β-carotene from 300 g pureed, cooked spinach, and 3 mg [3H]-vitamin A</td>
<td>21:1</td>
<td>19</td>
</tr>
<tr>
<td>[3H]-Labeled carrot, and vitamin A in oil capsule, n = 7</td>
<td>Comparison of AUC responses to the carrot and the vitamin A reference dose in adults</td>
<td>Calculated 11 mg trans-β-carotene from 100 g pureed, cooked carrot, and 3 mg [3H]-vitamin A</td>
<td>15:1</td>
<td>19</td>
</tr>
<tr>
<td>Fruit, n = 69; leafy vegetables, n = 70; retinol-rich foods, n = 70; control, n = 68</td>
<td>Changes of serum retinol concentration in lactating women after they ate fruit or vegetables, or took preformed vitamin A</td>
<td>Fruit: 4.8 mg trans-β-carotene; vegetables: 5.6 mg trans-β-carotene; retinol-rich diet: 610 μg retinol; control: 0.6 mg β-carotene and 1 μg retinol</td>
<td>12:1</td>
<td>17</td>
</tr>
<tr>
<td>Spirulina powder, n = 10</td>
<td>Comparison of AUC responses to the Spirulina and the vitamin A reference dose in adults</td>
<td>4.3 mg Spirulina trans-β-carotene</td>
<td>28:1</td>
<td>31</td>
</tr>
<tr>
<td>Golden Rice, n = 5</td>
<td>Comparison of AUC responses to a Golden Rice meal and the vitamin A reference dose in adults</td>
<td>Rice meal that contained 1–1.5 mg rice β-carotene</td>
<td>3.6:1</td>
<td>32</td>
</tr>
</tbody>
</table>

*RE, retinol equivalents; AUC, area under the curve.*